

## 2,6-Dansyl Azide as a Fluorescent Probe for Hydrogen Sulfide

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Received: 9 June 2013 / Accepted: 2 September 2013 / Published online: 1 October 2013  
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**Abstract** A second-generation sulfonyl azide-based fluorescent probe, 2,6-DNS-Az, has been developed for the quantitative detection of H<sub>2</sub>S in aqueous media such as phosphate buffer and bovine serum. Compare to the first-generation 1,5-DNS-Az probe, this probe shows both high sensitivity in phosphate buffer without the need for addition of surfactant and selectivity for sulfide over other anions and biomolecules, and thus can be used as a useful tool for detection of H<sub>2</sub>S in the biological system.

**Keywords** Hydrogen sulfide · Gasotransmitter · Fluorescent probe · Azide · Redox sensing

### Introduction

The past decade has seen a boost of research interest in hydrogen sulfide (H<sub>2</sub>S), which is recognized as one of the three important gasotransmitters including nitric oxide (NO) [1] and carbon monoxide (CO) [2]. H<sub>2</sub>S is synthesized endogenously and is involved in the regulations of a series of important genes. Endogenous concentrations of H<sub>2</sub>S is related to a number of diseases such as Down syndrome [3] and lung diseases [4]. H<sub>2</sub>S was also found to show protective effects in the cardiovascular (CV) [5] and central nervous systems (CNS) [6] and to play a regulatory role in inflammation [7,

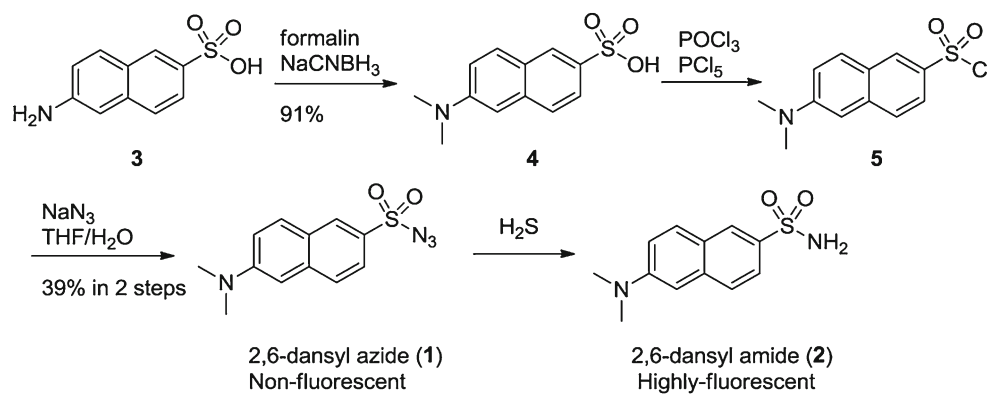
8]. Regulation of H<sub>2</sub>S levels is also a potential drug development strategy [9, 10]. The importance of accurate detection of H<sub>2</sub>S cannot be over-emphasized. However, this is not a trivial issue mainly due to the low stability of this gaseous molecule. H<sub>2</sub>S is volatile and prone to oxidation. Fluorescent probes are emerging as important tools for the selective and sensitive detection of H<sub>2</sub>S [11–13]. Chang and our group first reported that the reduction of an azido group attached to a fluorophore could be utilized for the design of H<sub>2</sub>S-selective fluorescent probes [14, 15]. Recently, a number of fluorescent probes have been developed, including probes based on nucleophilic cyclization [16–20], redox reactions [15, 21–24], and metal-sulfide formation [25–27]. A redox-based fluorescent probe (DNS-Az), which was reported by our group, shows significant fluorescence “turn-on” effect in the presence of H<sub>2</sub>S in aqueous solutions. Most importantly, the sensing reaction (completes in minutes) is the fastest among all redox-based fluorescent probes for H<sub>2</sub>S. This might be attributed to the unique structural features of DNS-Az. Compared to other redox-based fluorescent probes bearing an azido group, DNS-Az includes a sulfonyl azide. We believe that the sulfonyl azide provides the probe with fast reaction rates and increased photo-stability. However, the quantum yield of the fluorescent species DNS-NH<sub>2</sub> is very low (< 0.05) in pure aqueous solutions. In order to reach higher sensitivity, a surfactant Tween-20 was added in previous experiments [14, 28]. Herein we describe the synthesis and evaluation of a new fluorescent probe, which shares structure similarity with the previously reported probe but emits at a different wavelength with a higher fluorescence quantum yield and detection sensitivity in phosphate buffer without the need for addition of any surfactant.

The structure of DNS-Az is based on a 1,5-dansyl fluorophore, which is known for its large Stokes shift ( $\lambda_{\text{ex}}=330$  nm,  $\lambda_{\text{em}}=517$  nm in phosphate buffer/Tween-20). 2,6-Dansyl fluorophore has a smaller Stokes shift, however

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**Electronic supplementary material** The online version of this article (doi:10.1007/s10895-013-1296-5) contains supplementary material, which is available to authorized users.

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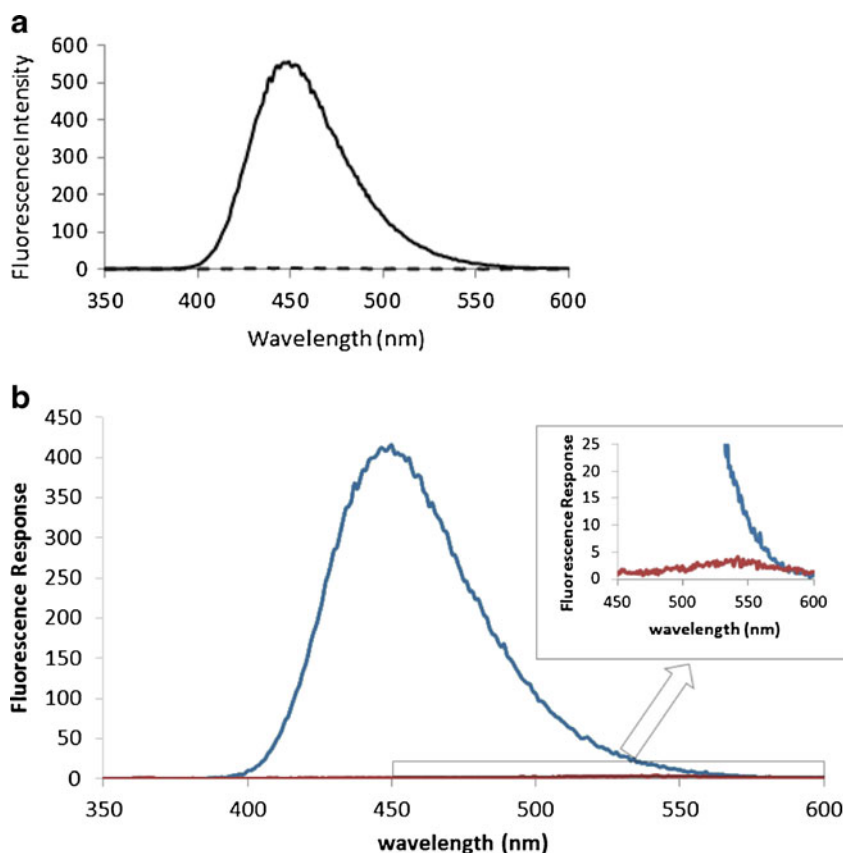
**Scheme 1** Synthesis of 2,6-dansyl azide

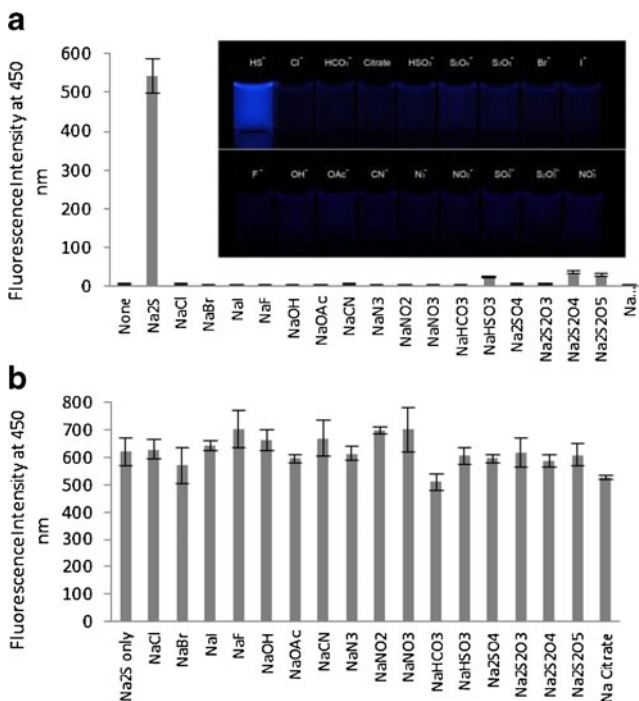
exhibits a much higher fluorescence quantum yield in pure aqueous media [28]. Therefore, it is conceivable that 2,6-dansyl azide (2,6-DNS-Az, **1**) might serve as a fluorescent probe for H<sub>2</sub>S with high detection sensitivity in pure aqueous media. Starting from commercially available 6-amino-2-naphthalenesulfonic acid (**3**), 2,6-dansyl azide was synthesized in 3 easy steps, including reductive alkylation to form dimethylamino sulfonate **4**, formation of sulfonyl chloride **5**, and substitution with sodium azide to afford the sulfonfyl azide **1** in 35 % overall isolated yield. (Scheme 1)

2,6-Dansyl azide was evaluated as a fluorescent probe for H<sub>2</sub>S. In all experiments, Na<sub>2</sub>S was used as the sulfide source in aqueous solutions as reported previously [14] and elsewhere.

As expected, 2,6-DNS-Az could be easily reduced to its corresponding sulfonamide after sulfide addition. 2,6-Dansyl amide **2** was isolated and identified using <sup>1</sup>H NMR, <sup>13</sup>C NMR and mass spectrometry (see supporting information). The quantum yield of 2,6-dansyl amide **2** was about 40-fold higher than that of 1,5-dansyl amide in deionized water (see supporting information). In a sulfide detection experiment, 20 μM of probe **1** showed an over 100-fold fluorescence increase in 60 min after addition of only 10 μM of sulfide in phosphate buffer without surfactant. This is a much more significant change compared to the 8-fold fluorescence increase observed for the first-generation probe 1,5-DNS-Az in phosphate buffer (Fig. 1b).

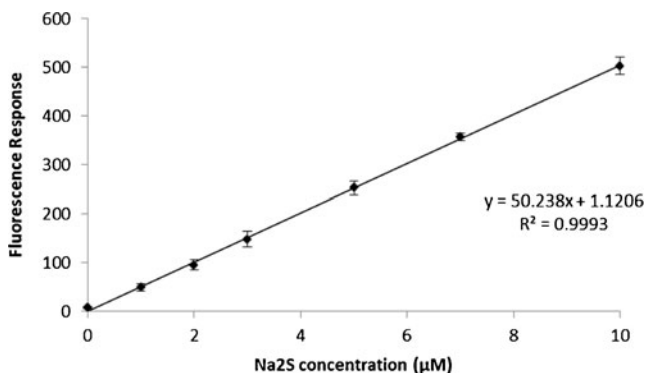
**Fig. 1** **a** Fluorescence increase of 2,6-DNS-Az (**1**, 20 μM) with the addition of sulfide (10 μM) in 0.1 M sodium phosphate buffer (pH 7.4) at room temperature. Dashed line represents fluorescence spectrum of **1** alone. Solid line represents fluorescence spectrum of **1** with sulfide. **b** Comparison of fluorescence response of 2,6-DNS-Az and 1,5-DNS-Az in phosphate buffer. Blue line represents fluorescence response of 2,6-DNS-Az (20 μM) to sulfide (10 μM); red line represents fluorescence response of 1,5-DNS-Az (20 μM) to sulfide (10 μM) in 0.1 M phosphate buffer (pH=7.4), λ<sub>ex</sub>=325 nm



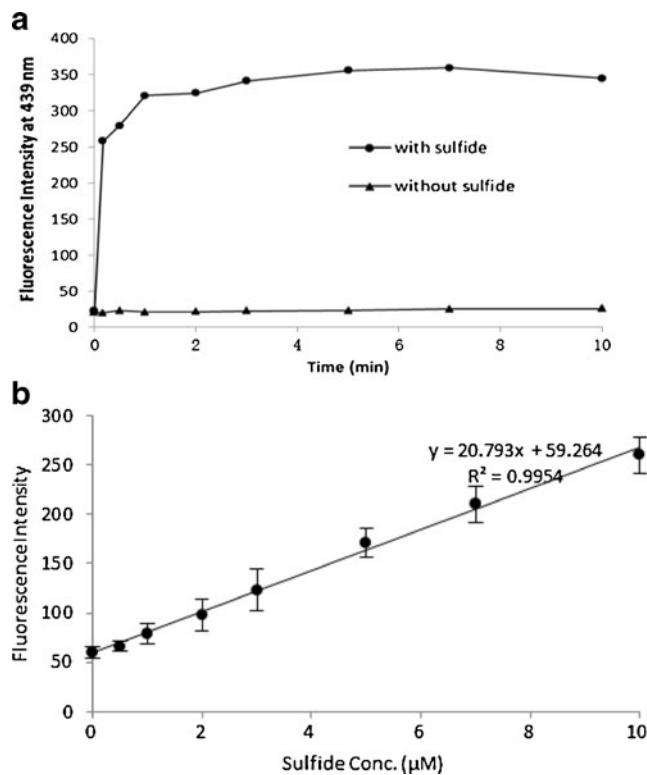


**Fig. 2** **a** Fluorescence changes of 2,6-DNS-Az (**1**) in the presence of various anions; **b** fluorescence changes of 2,6-DNS-Az (**1**, 20  $\mu$ M) in the presence of sulfide and various anions ( $\text{HS}^-$  10  $\mu$ M;  $\text{HSO}_3^-$ ,  $\text{S}_2\text{O}_4^{2-}$ , and  $\text{S}_2\text{O}_5^{2-}$  20  $\mu$ M;  $\text{Cl}^-$ ,  $\text{HCO}_3^-$ , citrate 100 mM all other anions 1 mM in 0.1 M sodium phosphate buffer (pH 7.4),  $\lambda_{\text{ex}}=325$  nm). Anions tested:  $\text{Cl}^-$ ,  $\text{Br}^-$ ,  $\text{I}^-$ ,  $\text{F}^-$ ,  $\text{OH}^-$ ,  $\text{OAc}^-$ ,  $\text{CN}^-$ ,  $\text{N}_3^-$ ,  $\text{NO}_2^-$ ,  $\text{NO}_3^-$ ,  $\text{HCO}_3^-$ ,  $\text{HSO}_3^-$ ,  $\text{SO}_4^{2-}$ ,  $\text{S}_2\text{O}_3^{2-}$ ,  $\text{S}_2\text{O}_4^{2-}$ ,  $\text{S}_2\text{O}_5^{2-}$ , and citrate. Data represents average of three independent experiments

After the fluorescence response of 2,6-DNS-Az to sulfide was demonstrated, a variety of 17 common anions, including sulfur-containing reducing anions, were used at concentrations above their physiological levels to test the selectivity of 2,6-DNS-Az. It was found that 2,6-dansyl azide showed almost exclusive response to sulfide over other anions (Fig. 2). For strong reducing anions such as  $\text{HSO}_3^-$ ,  $\text{S}_2\text{O}_4^{2-}$ , and  $\text{S}_2\text{O}_5^{2-}$ , the selectivity was found to be 30 to 44-fold. Other reducing biological species, such as cysteine (Cys) and glutathione



**Fig. 3** Hydrogen sulfide concentration dependent fluorescence intensity changes: 2,6-dansyl azide 20  $\mu$ M,  $\text{Na}_2\text{S}$  0–10  $\mu$ M in 100 mM phosphate buffer (pH 7.4,  $\lambda_{\text{ex}}=325$  nm), Data represents average of three independent experiments



**Fig. 4** **a** Reaction time profile of 2,6-DNS-Az (**1**, 20  $\mu$ M) and  $\text{H}_2\text{S}$  (10  $\mu$ M) in FBS; **b**  $\text{H}_2\text{S}$  concentration dependent fluorescence intensity changes, determined using a fluorometer: 2,6-DNS-Az 20  $\mu$ M,  $\text{Na}_2\text{S}$  0–10  $\mu$ M in FBS ( $\lambda_{\text{ex}}=325$  nm). Concentration dependence data represents average of three independent experiments

(GSH) at 100  $\mu$ M, were also tested (Figure S3). The probe did not show fluorescence response to Cys and GSH. A slight increase in fluorescence response to sulfide was found in the presence of Cys and GSH. This could be due to antioxidative effect of Cys and GSH, which prevented undesired oxidation of sulfide in the reaction media. Selectivity over nucleophilic species was also tested by using high concentrations (1 mM) of glycine and lysine. These results have indicated that 2,6-DNS-Az shows sufficient selectivity for the detection of  $\text{H}_2\text{S}$  in a complex biological sample.

For all quantitative analytical methods, a linear calibration curve is always desired because it allows easy calculation. After confirming the selectivity of 2,6-DNS-Az, this probe was evaluated for quantitative measurement of  $\text{H}_2\text{S}$  by testing concentration-dependent response of 2,6-DNS-Az **1** to sulfide. As shown in Fig. 3, a linear correlation ( $R^2=0.9993$ ) was found for sulfide in phosphate buffer. As determined by 3:1 signal/noise ratio, the detection limit for sulfide in phosphate buffer is 1  $\mu$ M (Figure S4). This indicates that 2,6-DNS-Az could be used for quantitative measurement of  $\text{H}_2\text{S}$  in pure aqueous media.

Due to the biological significance of  $\text{H}_2\text{S}$ , its detection in biological systems such as blood serum is of great importance in both research and clinical applications. Therefore, we are also interested in using 2,6-DNS-Az for sulfide detection in

serum. The probe was evaluated in fetal bovine serum (FBS). First, a reaction time profile was tested for 2,6-DNS-Az and sulfide. As shown in Fig. 4, although FBS shows some background fluorescence, a very significant increase was still observed after addition of sulfide. Over 10-fold fluorescence increase was observed for only 10  $\mu\text{M}$  of sulfide. This is more than 2-fold higher sensitivity compared to 1,5-DNS-Az reported earlier by our group. The reaction of 2,6-DNS-Az with sulfide is very fast and completes in about 2 min in FBS. This is very important considering that  $\text{H}_2\text{S}$  is very unstable and easily consumed in biological systems. We also tested concentration-dependent fluorescence changes of 2,6-DNS-Az in FBS. The linear calibration curve ( $R^2=0.995$ ) obtained in FBS has also indicated that 2,6-DNS-Az could serve as a useful tool for  $\text{H}_2\text{S}$  detection. The detection limit of  $\text{H}_2\text{S}$  in FBS is about 6  $\mu\text{M}$ .

## Conclusions

The biological significance as well as difficulty in accurate detection of  $\text{H}_2\text{S}$  demonstrates the importance of finding new detection methods. We have reported previously a fluorescent probe (1,5-DNS-Az) for  $\text{H}_2\text{S}$  detection, which requires the addition of Tween-20 to achieve sufficient sensitivity when used in buffer solutions. The present work reports the development and evaluation of a new fluorescent probe for  $\text{H}_2\text{S}$ , 2,6-DNS-Az. This probe shows much higher fluorescence change in pure aqueous media (>100-fold for 10  $\mu\text{M}$  of sulfide) compared to the probe described earlier (8-fold) without addition of any surfactant. High detection sensitivity, almost exclusive selectivity and excellent linear correlation for sulfide in aqueous solution and blood serum make this probe a useful tool for quantitative detection of  $\text{H}_2\text{S}$ .

**Acknowledgment** We are grateful for the financial support from Chinese Scholarship Council (KW), Molecular Basis of Disease for a fellowship (HP), and Center for Diagnostics and Therapeutics for a CDT/University fellowship (HP).

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